



Providing innovative strategies for high content imaging and analysis

Patrick M. McDonough, Ph.D. VP of Biology, Vala Sciences Inc Computational Toxicology Communities of Practice webinar Nov. 17, 2011

A brief history of Vala Sciences Inc:

The Vala Sciences Inc was founded in 2003. The biology and engineering team previously founded Q3DM (Quantitative 3-Dimensional Microscopy), a San Diego based company that developed the EIDAQ/IC-100 one of the first High Content Assay workstations.

Jeffrey Price, MD. Ph.D., the CEO of Vala, and cofounder of both Q3DM and Vala is a pioneer in the field of High Content Analysis (HCA, typically cell-based assays which feature high resolution, multi-channel fluorescence imaging, with automated image acquistion and image analysis software).

Vala Sciences offers instrumentation, software, reagents, and assays for HCA for both cells and tissue, relevant to high throughput screening, preclinical research, and digital pathology applications.

Recent funded grants and contracts to Vala Sciences

Grants:

- 2006 NIH SBIR 1R43DK074333-01 Title: *HT Image Assay of Lipid Droplet Formation in Human Adipocytes*. Phase I = \$142,606, Phase II = \$1,503,000. PI: McDonough
- 2006 NIH STTR 1R41DK076510-01 Title: *Development automated assay-regulators insulin synthesis*. Phase I = \$200,371. PI: McDonough
- 2006 NIH FastTrack STTR 1R42HL086076-01 Proposal Title: Live cell and HCS assays to quantify production of cardiomyocytes from stem cells. Phase I = \$249,866, Phase II = \$1,305,000. PI: McDonough
- 2009 NIH 1R03MH082378-01 Proposal Title: High Throughput Imaging Assay for Beta-Catenin. \$25,000 (NIH Network Molecular Libraries Probe Production Centers). PI: McDonough
- 2009 NIH 1R03DA026213-01 Proposal Title: A High Throughput Imaging Assay for Hepatic Lipid Droplet Formation. \$25,000 (NIH Network Molecular Libraries Probe Production Centers. PI: McDonough
- 2009 CIRM Title *Differentiation of ventricular, atrial, and pacemaker type cardiomyocytes from stem cells*. 2 years, \$900,000. PI: McDonough
- 2010 NIH SBIR phase I Title: *High content analysis of mitochondrial replication* Phase I = \$270,000 PI: Whittaker
- 2010 NIH STTR 1R41DK082087-01 Title Automated quantification of lipid droplets in fatty liver tissue sections. Phase I = \$298,466. Phase II budget = \$1,464,024. PI: McDonough
- 2010 NIH STTR 1R41AR055604-01A2 Title Automated analysis of skeletal muscle fiber crossectional area and metabolic type. Phase I = \$153,853. Phase II budget = \$1,200,000. PI: McDonough
- 2011 (nearly approved) NIH STTR FastTrack Title: *Optogenetic Multiparametric Assay for HT Cardiotoxicity Testing*. PI: Cerignoli.

Contracts:

- 2009 NIH FastTrack SBIR contract Title: *Drug Safety Assessment in iPS Derived Cardiomyocytes*. Phase I = \$119,000 Phase II = \$840,000. PI: Cerignoli
- 2010 NIH SBIR FastTrack contract Title: *Hapten and Qdot based assay for breast cancer biomarkers*. Phase I = \$150,000 Phase II = \$1,700,000. PI: McDonough
- 2011 NIH SBIR contract Title: Automated karyometry as a companion diagnostic for chemoprevention of breast cancer Phase I = \$204,000 PI: McDonough

Overall, Vala has received approx. \$12,500,000 in funding from grants and contracts (the NIH has been the primary source of grant/contract support).

Additional recent contracts include:

Johnson & Johnson - project to develop Kinetic Image Cytometry (KIC) methods for simultaneous measurement of intracellular calcium and voltage in cardiac myocytes.

Sanofi Aventis - project to test candidate pharmaceuticals for effects on voltage-dependent channels in cardiac myocytes using KIC

Vala has also recently been approved for a \$1,500,000 loan from the SBA (Silvergate Bank, San Diego, CA).

Vala Collaborators:

Sanford-Burnham Medical Research Institute.

Sanford-Burnham has previously collaborated with Vala Sciences Inc on the following grants and contracts including:

STTR Fast-Track 1 R42 HL086076-01 "Live cell and HCS assays to quantify production of cardiomyocytes from stem cells" and SBIR contract HHSN268200900044C "Drug Safety Assessment in IPS Derived Cardiomyocytes").

Two of Vala's MLSCN/MLPCN assays (to identify activators of beta-catenin (R03 MH082378), and inhibitors of hepatic lipid droplet formation (R03 MH083261) were screened on large chemical compound libraries (approx. 200,000 compounds) by the Conrad Prebys Center for Chemical Genomics at Sanford-Burnham.



Sanford Burnham Medical Research Institute From Research, The Power To Cure

Conrad Prebys Center Chemical Genomics

Conrad Prebys Center for Chemical Genomics

One of the most advanced infrastructure for small molecule drug discovery in non-profit world.

Staffed by ~80 professionals, most with pharmaceutical company experience
 Functional Units include:

- Assay Development (including advanced HC
- Assay Development (including advanced HCS for phenotypic screens)
 HTS & Compound library management (including ultra-HTS robotics and total libraries >
 900K compounds)
- Chemical fragment screening by NMR
- Affinity Selection-Mass Spectrometry (ASMS) screening
- Cheminformatics
- Medicinal Chemistry
- Pharmacology
- Structure-based drug optimization (robotic protein crystallography) (NMR)
- Current throughput is ~40 HTS campaigns per year, current capacity to undertake ~50/year.

World Class HTS Capabilities





Instrumentation:

2003 Q3DM Eidaq IC100 (also marketed by Beckman Coutler) (rapid autofocus, multichannel fluorescence, microtiter plates, slides)

Present: IC200 and IC300 product line. Even faster autofocus, continuous scanning, large format cameras, Kinetic Image Cytometry

Recent customers include the Sanford Burnham Medical Research Institute, University of Houston, Baylor College of Medicine, Genomics Institute of the Novartis Research Foundation.



Jeffrey H. Price, M.D., Ph.D.



Autofocus based upon chromatic abberation

Prosstate biopsy visualized via fluorescence microscopy

Software: CyteSeer®

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Randall Ingermanson



Jeff Hilton

Features of CyteSeer®

- Coded in Java, cross-platform compatible
- True cell-by-cell cytometry
- Plug-in and pipe-line architecture.
- Easily modified and optimized for specific applications
- Can be downloaded and tested for free from our website

http://www.valasciences.com





The Lipid Droplet/Colocalization Algorithm:

(human adipocytes, lipolysis assay (nuclei, lipid droplets, phosphoperilipin)





A rich set of data parameters are derived by CyteSeer for each cell.

Membrane algorithm (94) Lipid droplet/colocalization algorithm (98)

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| F08 | Gates | 3 F01 0 0 36.4664 | 23.9504 22.1885 | 19.7820 | 22.1360 115.8369 | 33.0744 255.0000 2 | 55.0000 255.0000 | 255.0000 | 243.6000 | 255.0000 1929.000 | |
| Well F09 | Source Gate | 4 F01 0 0 43.9259 | 32.6311 14.4042 | 14.0121 | 23.2203 123.2816 | 19.1840 38.1118 | 93.7778 88.8738 | 93.5106 38.071 | 9 132.7647 | 39.0616 5402.000 | |
| F10 | CyteSeer Remove Two Percent | 5 F01 0 0 54.2505 | 28.9812 37.2701 | 34.4580 | 27.7613 102.2895 | 27.3500 53.5185 | 58.3111 53.3120 | 63.6500 53.280 | 1 53.2689 | 53.4888 2561.000 | |
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| F13 | CyteSeer Remove Boundary C | 7 F01 0 0 57.0148 | 27.6432 27.6494 | 24.2805 | 25.9666 90.8865 | 36.0880 45.9401 | 56.6946 46.8811 | 64.7805 44.58 | 1 46.7339 | 46.0992 1770.000 | |
| F14 4 | CyteSeer Column A | 8 F01 0 0 37.0526 | 15.3393 16.4457 | 16.1412 | 15.3393 91.9013 | 43.1806 235.5116 1 | 60.6667 255.0000 | 245.4131 111.892 | 5 | 240.1869 1806.00(| |
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| Clear All Select All | New Edit Remove | 13 F01 0 0 32.1225 | 14.3642 13.8857 | 13.8371 | 14.2956 92.7415 | 16.3743 31.5181 | 38.6716 36.3566 | 51.3084 31.452 | 6 35.0665 | 31.5496 81051.00 | |
| | Data is: Ungated Gated | 14 F01 0 0 44.9502 | 16.5046 19.5410 | 17.9650 | 16.1432 137.9032 | 35.0012 216.7628 1 | 52.0093 252.3848 | 241.3411 88.04 | 2 | 219.8424 4586.000 | |
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| | Significant Figures: | 16 F01 0 0 20.3333 | 13.9526 14.1613 | 14.1552 | 13.9526 54.2414 | 27 5688 34 9054 | 40.7083 67.6853 | 207.0974 45.49 | 0 65.4165 | 82.1240 8383.000 | |
| Gates | | 18 F01 0 0 65 9076 | 41 7283 41 5497 | 37 0406 | 28 1373 120 7658 | 39 0906 56 2682 | 50.8022 56.7049 | 68 9844 55 62 | 3 56 3621 | 56 2928 2394 000 | |
| | Cell ID Count: 212/212 [100 %] | 19 F01 0 0 65.4069 | 34.2299 44.4913 | 39.9748 | 28.3782 112.6090 | 39.0621 101.6786 | 95.1219 61.5948 | 198.2210 62.323 | 4 61.5948 | 95.4594 2172.000 | |
| Name Source DataTable | | 20 F01 0 0 27.0765 | 15.8356 16.2871 | 16.0280 | 15.8356 121.2012 | 31.5502 249.9206 2 | 55.0000 255.0000 | 252.2285 92.909 | 1 | 250.2500 7455.000 | |
| | All Measurements | 21 F01 0 0 62.7562 | 14.1529 15.2135 | 14.4615 | 14.1529 96.9253 | 24.6247 96.6584 | 99.3760 62.5896 | 224.4894 50.883 | 4 60.7669 | 94.0864 13936.00 | |
| | | 22 F01 0 0 47.5876 | 18.1913 21.2273 | 19.8907 | 17.6851 118.3739 | 26.2258 54.7783 | 96.0458 48.2804 | 196.9057 46.702 | 9 48.2804 | 54.3495 6510.000 | |
| | * | 23 F01 0 0 | 15.6862 14.3302 | 14.2876 | 15.6862 78.7973 | 18.5982 40.4159 | 78.2534 | 94.0333 40.105 | 1 68.9931 | 41.5660 32728.00 | |
| | Measurement Show | 24 F01 0 0 32.9620 | 16 9904 17 1845 | 15.9029 | 16.5753 128.0018 | 42.2079 255.0000 2 | 55.0000 255.0000 | 253.0000 | 2 | 255.0000 4045.000 | |
| | Name of the well | 26 F01 0 0 35.4589 | 22.6409 14.3588 | 14.1331 | 20.5586 152.3149 | 18.0746 36.2775 | 75.5308 62.6547 | 77.9798 36.03 | 6 60.6456 | 36,5369 36450.00 | |
| | Index of the z-slice | 27 F01 0 0 34.6396 | 15.7267 17.1986 | 16.6799 | 15.6539 129.2574 | 27.2655 150.3983 1 | 77.2792 251.6238 | 221.5924 71.356 | 0 | 156.1974 8310.000 | |
| | Average Pixel Intensity of Lini | 28 F01 0 0 53.3257 | 28.4767 25.0970 | 22.5412 | 27.1962 104.7312 | 28.2902 40.3707 | 49.5029 45.0573 | 55.1667 40.242 | 4 44.7000 | 40.9145 2401.000 | |
| | Average Pixel Intensity of Lipi | 29 F01 0 0 35.6103 | 16.1287 16.9276 | 16.2310 | 15.8357 145.0992 | 28.4175 36.2139 | 43.5897 40.9544 | 51.1964 36.038 | 1 40.9544 | 36.5776 4830.000 | |
| | Average Pixel Intensity of Lipi | 30 F01 0 0 22.8788 | 15.0593 14.6891 | 14.6412 | 15.0593 98.5826 | 21.6757 41.3935 | 48.1818 46.0996 | 41.393 | 5 46.0996 | 41.6205 9313.000 | |
| • | Average Pixel Intensity of Lipi | 31 F01 0 0 26.7429 | 17.2671 15.8972 | 15.3857 | 16.9498 165.0662 | 32.4591 34.9868 | 41.1214 41.3539 | 44.7143 34.934 | 6 41.3539 | 35.5916 3937.000 | |
| | Average Pixel Intensity of Lipi | 33 F01 0 0 33 9813 | 17.7101 10.2847 | 15.5550 | 15 1938 79 5046 | 23 1374 37 7787 | 45 5875 34 7587 | 49 5946 37 62 | 2 34 7587 | 37 5108 5839 000 | |
| 1 - | Average Pixel Intensity of Nu | 34 F01 0 0 71.9276 | 32,1496 39.6290 | 32,7992 | 30.0785 119.5536 | 60.5399 53.7699 | 61.5882 57.0224 | 66.0816 53.054 | 4 56,7700 | 55.0826 678.000 | |
| | Average Pixel Intensity of Pro | 35 F01 0 0 68.0451 | 28.5448 33.2239 | 27.9283 | 28.3066 129.7534 | 35.0564 50.8450 | 61.0855 56.6345 | 66.9753 50.450 | 8 56.2668 | 51.5851 2800.000 | |
| | Average Pixel Intensity of Pro | 36 F01 0 0 100.5865 | 27.3629 19.3371 | 14.9803 | 20.6487 72.5327 | 18.8976 33.1986 | 67.4663 41.1480 | 75.4697 32.903 | 5 38.9457 | 33.5537 13553.00 | |
| | Average Pixel Intensity of Pro | 37 F01 0 0 26.6063 | 14.8088 13.9127 | 13.8720 | 14.7114 153.7328 | 18.1494 37.0237 | 41.0079 46.4865 | 56.8077 36.963 | 0 46.3337 | 37.1609 55465.00 | |
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Data parameters related to intensity of different cellular compartments in different image channels

| TII Mi Nm | Total Integrated Intensity of Membrane Image on Nucleus Mask |
|-----------|--|
| API MI NM | Average Pixel Intensity of Membrane Image on Nucleus Mask |
| MPI Mi Nm | Median Pixel Intensity of Membrane Image on Nucleus Mask |
| SPI Mi Nm | Standard Deviation of Pixel Intensity of Membrane Image on Nucleus Mask |
| TII Mi Mm | Total Integrated Intensity of Membrane Image on Membrane Mask |
| API Mi Mm | Average Pixel Intensity of Membrane Image on Membrane Mask |
| MPI Mi Mm | Median Pixel Intensity of Membrane Image on Membrane Mask |
| SPI Mi Mm | Standard Deviation of Pixel Intensity of Membrane Image on Membrane Mask |
| TII Mi Cm | Total Integrated Intensity of Membrane Image on Cytoplasm Mask |
| API Mi Cm | Average Pixel Intensity of Membrane Image on Cytoplasm Mask |
| MPI Mi Cm | Median Pixel Intensity of Membrane Image on Cytoplasm Mask |
| SPI Mi Cm | Standard Deviation of Pixel Intensity of Membrane Image on Cytoplasm Mask |
| TII Mi Wm | Total Integrated Intensity of Membrane Image on Whole-Cell Mask |
| API Mi Wm | Average Pixel Intensity of Membrane Image on Whole-Cell Mask |
| MPI Mi Wm | Median Pixel Intensity of Membrane Image on Whole-Cell Mask |
| SPI Mi Wm | Standard Deviation of Pixel Intensity of Membrane Image on Whole-Cell Mask |
| | |

Colocalization of labels between differerent

- .
- image channels for different cellular compartments

PCC Ni Mi Wm Pearson Correlation Coefficient of Nucleus Image vs Membrane Image on Whole-Cell Mask K1 Ni Mi Wm Manders K1 Coefficient of Nucleus Image vs Membrane Image on Whole-Cell Mask K2 Ni Mi Wm Manders K2 Coefficient of Nucleus Image vs Membrane Image on Whole-Cell Mask Manders Overlap Coefficient of Nucleus Image vs Membrane Image on Whole-Cell Mask MOC Ni Mi Wm M1 Ni Mi Wm Manders M1 Coefficient of Nucleus Image vs Membrane Image on Whole-Cell Mask M2 Ni Mi Wm Manders M2 Coefficient of Nucleus Image vs Membrane Image on Whole-Cell Mask Masked PCC Ni Mi Nm Masked Pearson Correlation Coefficient of Nucleus Image vs Membrane Image on Nucleus Mask Masked K1 Ni Mi Nm Masked Manders K1 Coefficient of Nucleus Image vs Membrane Image on Nucleus Mask Masked K2 Ni Mi Nm Masked Manders K2 Coefficient of Nucleus Image vs Membrane Image on Nucleus Mask Masked MOC Ni Mi Nm Masked Manders Overlap Coefficient of Nucleus Image vs Membrane Image on Nucleus Mask Masked M1 Ni Mi Nm Masked Manders M1 Coefficient of Nucleus Image vs Membrane Image on Nucleus Mask Masked M2 Ni Mi Nm Masked Manders M2 Coefficient of Nucleus Image vs Membrane Image on Nucleus Mask

Kinetic Image Cytometer (KIC)



Fabio Cerignoli



Ross Whittaker

















Data set #1



Data set #2



Data set #3



Vala is offering the following assays related to KIC-Cardiac Myocyte:

CD1, Cardiac differentiation , human cardiac myocytes (long term effects on iPS-derived cardiac myocytes)

CT1, Cardiac toxicity, human cardiac myocytes (acute effects on iPS-derived cardiac myocytes)

Adipogenesis

Excess adipogenesis, leads to obesity. Abnormally low adipogenesis may contribute to cachexia.

2006 NIH SBIR 1R43DK074333-01 Title: *HT Image Assay of Lipid Droplet Formation in Human Adipocytes*.

Collaborators: Zen-Bio Inc (suppliers of primary adipocytes and other cell types), Baylor College of Medicine (development of custom antibodies)

The basics of assay development were published in a peer-reviewed paper:

McDonough, P. M., Agustin, R. M., Ingermanson, R. S., Loy, P. A., Buehrer, B. M., Nicoll, J. B., Prigozhina, N. L., Mikic, I., Price, J. H. 2009. Quantification of lipid droplets and associated proteins in cellular models of obesity via high content/ high throughput microscopy and automated image analysis. *ASSAY and Drug Development Technologies, 7:440-460*. NIHMSID#194280

Early experiment demonstrating the effect of rosiglitazone on differentiation of preadipocytes (A) to adipocytes (B), lipid droplets as a gray scale image (C), creation of the Lipid Droplet Mask (D), and the correlation between biochemical analysis of triglycerides (E) and lipid droplets as assayed via microscopy (F).





Data from the adipogenesis assay



Vala is offering the following assays related to Adipogenesis:

A1, Adipogenesis, Human adipocytes, agonists

A2, Adipogenesis, Human adipocytes, antagonists

Related assay:

LD2, Lipid droplet formation, Huh-7 cells (hepatocytes). This is related to fatty liver disease.

Lipolysis

McDonough, P. M., Ingermanson, R. S., Loy, P.A., Koon, E. D., Whittaker, R., Laris, C. A., Hilton, J. M., Nicoll, J. B., Buehrer, B. M., Price, J. H. (2010). Quantification of Hormone Sensitive Lipase Phosphorylation and Colocalization with Lipid Droplets in Murine 3T3L1 and Human Subcutaneous Adipocytes via Automated Digital Microscopy and High-Content Analysis. Assay Drug Dev Technol. 2010 Dec 27.



For lipid droplets to be metabolized, the process of lipolysis must be initiated.

The activation of lipolysis is associated with phosphorylation of perilipin (Peri), and Hormone Sensitive Lipase (HSL). Adipocyte triglyceride lipase (ATGL) is also critically important. Phosphorylation of perilipin attracts HSL and ATGL to the lipid droplets. HSL and perilipin are substrates for cAMP activated protein kinase (PKA), and cGMP-activated protein kinase (PKG)











Results from Vala's HCA-based lipolysis assay:



Vala is offering the following assays related to Lipolysis:

L1, Lipolysis, Human adipocytes, agonists

- L2, Lipolysis, Human adipocytes, antagonists
- L3, Lipolysis, murine 3T3L1 adipocytes, agonists

Cadherins and beta-catenin

Prigozhina, N. L., Zhong, L., Hunter, E. A., Mikic, I., Callaway, S., Roop, D. R., Mancini, M. A., Zacharias, D., Price, J. H., McDonough, P. M. (2007) Plasma membrane assays and three-compartment image cytometry for high content screening. ASSAY and Drug Development Technologies, 5:29-48



E-Cadherin



N-Cadherin



Beta-Catenin

A. Control



7.5 μM MSC#0267313



Vala is offering the following assays for Cadherins and Beta-Catenin:

ECAD1, E-Cadherin, A431 cells, agonists

NCAD1, N-Cadherin, HeLa cells, agonists

VCAD1, VE-Cadherin, Human vascular endothelial cells, agonists

BC1, Beta-Catenin, HeLa, agonists



Ross Whittaker

Assessing mitochondrial membrane potential ($\Delta \psi_M$) using HCA





Vala is offering the following assays related to Mitochondrial Function

MP1, Mitochondrial membrane potential, Huh-7 cells, acute

MP2 Mitochondrial membrane potential, Huh-7 cells, long term



Stem cell pluripotency

Mark Mercola

Assay SCP1, Stem cell pluripotency: Compounds that reduce pluripotency reduce GFP expression in human ES cells expressing GFP from an Oct4 promoter





Mesodermendoderm formation

Mark Mercola

Assay MEF1, Mesoderm endoderm formation, mouse embryonic stem cells

mESCs engineered to express eGFP under control of the vascular growth factor receptor-2. Cells are treated with compounds and assayed for both GFP (indicates mesoderm formation), and FoxA2 (indicates endoderm formation.







Heart cell formation

Mark Mercola

Assay HCF1: Heart cell formation, mouse embryonic stem cells

mESCs engineered to express eGFP under control of the alpha-myosin heavy chain promoter. Cells are subjected to a differentiation protocol in the presence of test compounds. Increases in GFP indicated increased formation of cardiac myocytes.

A related assay was recently published:

Willems, E., et al. 2011. Small-molecule inhibitors of the Wnt pathway potently promote cardiomyocytes from human embrynoic stem cellderived mesoderm. Circulation Ressarch 109:360-3641, Mesoderm endoderm formation, mouse embryonic stem cells



Pancreatic beta cell differentiation/ maturation

Pam Itkin-Ansari

Assay PBCD1: Pancreatic Beta Cell Differentiation

TPNE cells (derived from human fetal islets) express GFP downstream from the human insulin promoter. Compounds that promote differentiation of this pancreatic beta cell precursor towards the mature beta cell increase GFP expression.



Kiselyuk, A., et al. Phenothiazine neuroleptics signal to the human insulin promoter as revealed by a novel high-throughput screen. J Biomol Screen 15, 663-670 (2010).



The Notch pathway

Malene Hansen

The appearance of germ cells in C. elegans is dependent Notch activity and germ cells can be quantified via high content microscopy.

Assay CE1: C. elegans, Notch antagonists. Test for the ability of compounds to inhibit worms with a gain of function mutation in Notch.

Assay CE2: C. elegans, Notch agonists. Tests for the ability of compounds to increase germ cells in wild-type worms.



Alexi Terskikh

Assay NO1: Optogenetic-based assay of synaptic connectivity of human neurons

iPS- or neuronal-precursor-derived neurons are engineered to express channel rhodopsin, and loaded with fluo-4 to monitor calcium with KIC. Elevations in calcium in post-synaptic cells correspond to neurotransmission.

Assay MCDO1: Multiple choice differentiation outcomes

Utilizes pluripotent human neural crest stem cells. Quantifies the effects of compounds to influence differentiation to neurons, smooth muscle, glia, and melanocytes.

Vala people:

Andrew Heisel

Fabio Cerignoli

Ramses Agustin

Claire Weston

Robyn Garcia

James Evans

Ross Whittaker

Piyush Gehalot

Mike Markoudakis

Emily Arsenault

Constance Allison

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