

# RASL-Seq: A Gene Expression Platform to Identify Toxicity Mechanisms and Adaptive Responses

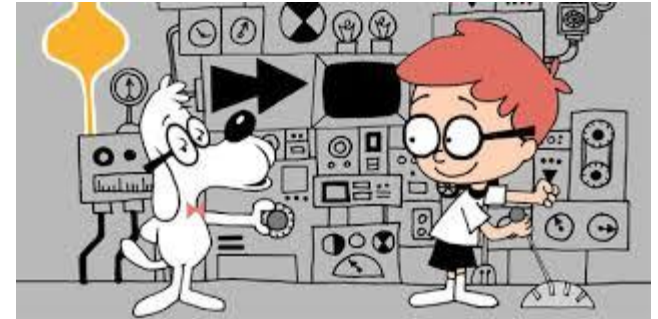
DAVID GERHOLD  
NIH - NCATS  
20 MARCH, 2014

EPA'S COMMUNITIES OF PRACTICE

NCATS

# Summary

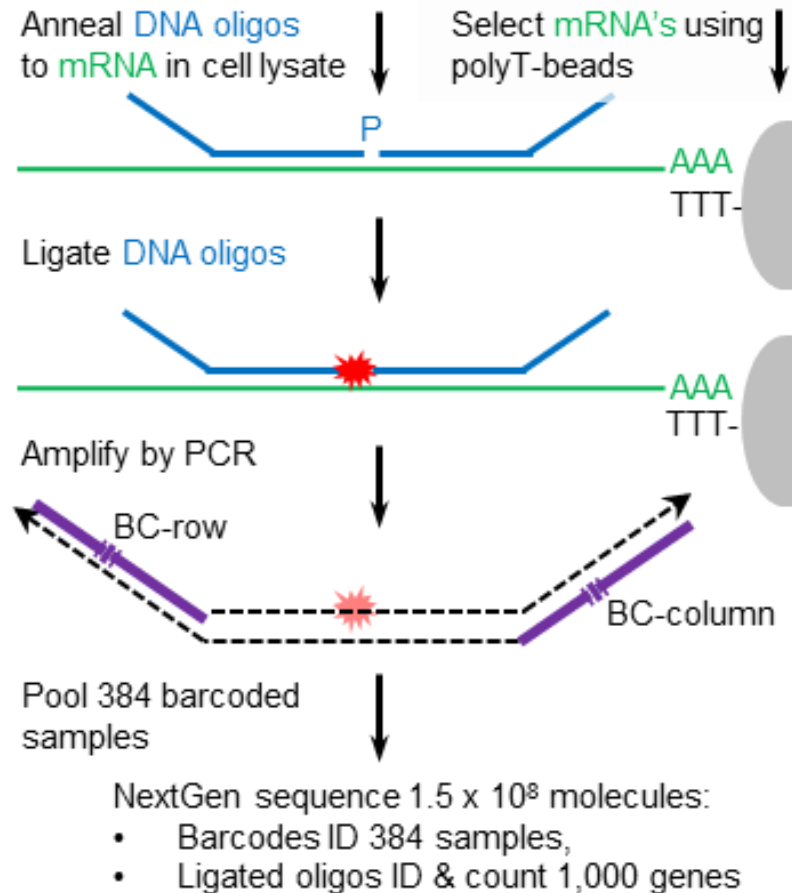
- RASL-Seq Selected, Results
- RASL-Seq platform works well
- Industrialization proceeding
- Selecting 1,400 genes to represent the genome(s)
- Goal: HT Gene Expression Core Facility
- How to apply it? Elucidate modes and mechanisms of toxicities or AEs. ‘Secondary screening’
- How will we use these results for risk assessment?



# Goals

- A technology that will quantify mRNA responses in hundreds-thousands of genes
- Throughput > Thousands of samples to address: many compounds x multiple cell lines x multiple concentrations x multiple time points
- >> Gene Expression Core Facility at NCATS
- Low variance, intra- & inter-experiment, will enable data interpretation and construction of a rich reference database
- Data analysis pipeline

# RASL-Seq Schematic



# RASL-Seq Characteristics

RASL-Seq has emerged from evaluation of six technologies based on:

- **Multiplex:**  $\geq 1,000$  genes/sample, and 384 samples/sequencing reaction
- **Throughput:** (384 samples/run x  $\sim 10$  runs/week)
- **Economy:** roughly \$10.43/sample or \$4,000/384 sample-run, including NextGen sequencing
- **Accuracy:** uses 3 redundant assays/gene
- **Reproducible:** avoids chaotic cDNA synthesis step. Intra-experiment  $R^2 \geq 0.99$
- **Gene-Specific:** % mispairing events (after removing 5 promiscuous assays, of 360) is 0.26%

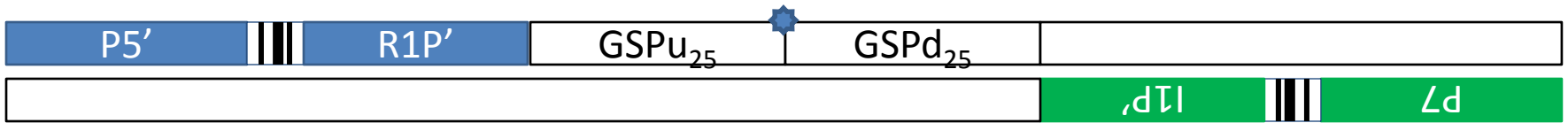
RASL-Seq invented by Xiang-Dong Fu Lab, Stanford U. Li H et al. PNAS 2012;109:4609-4614

Collaboration between BioSpyder (J. Yeakley & J. McComb) and NCATS to optimize RASL-Seq. Includes SBIR grant from NCATS.

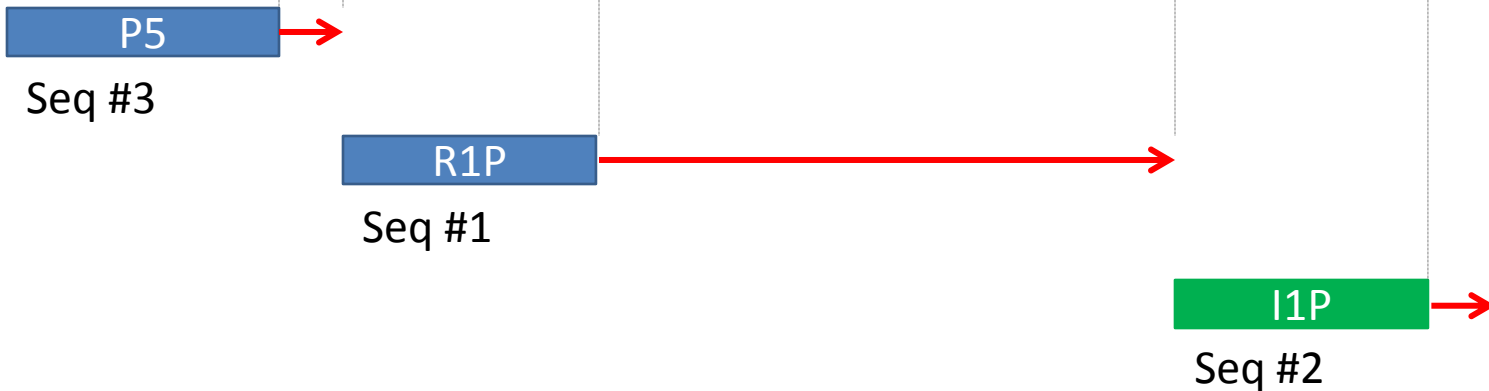


# RASL Product Sequencing

P

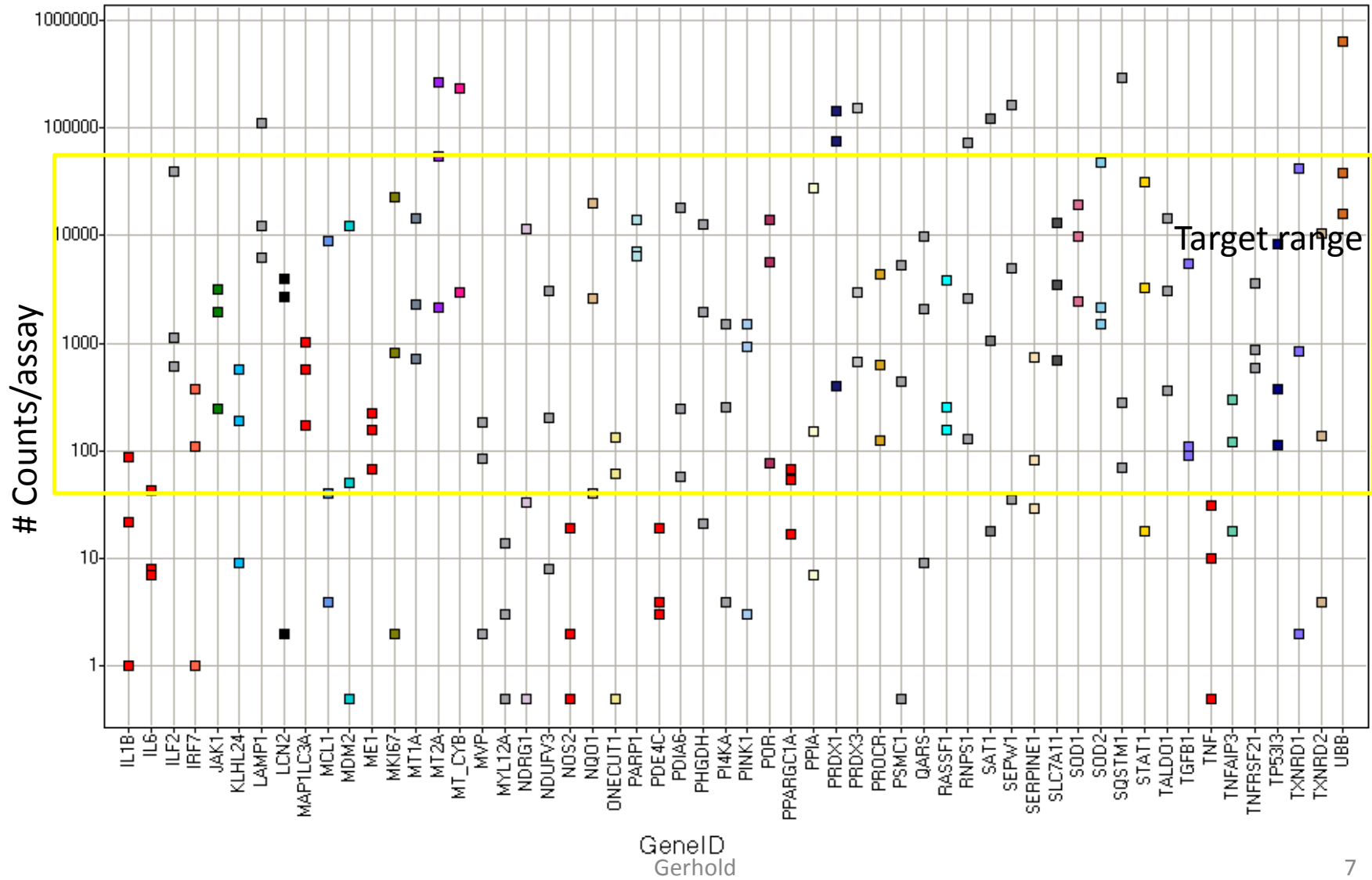


Illumina HiSeq Sequencing Reactions:



# 3 RASL-Seq assays/gene vary widely in # counts

- ⇒ Remove 'greedy' assays
- ⇒ ignore assays that give low counts



# RASL-Seq Progress March 2014

1<sup>st</sup> Gene set 120 genes (x 3 assays/gene) =360 assays. 13 'greedy' assays removed, and 5 non-specific (mispairing) assays removed.

- All steps performed by hand, using 384 samples, in microplates.
- Biomek FX robot installed 18 March 2014
- Good yields from PCR step in all five 'runs'
- 1-of-3 sequencing primers has been problematic. Two datasets now have resolved that issue for MiSeq instrument (24 or 50 samples  $\Rightarrow$  11 million high-quality seq). Excellent data!
- Testing on HiSeq2000 instrument set for April 2014. Expect 150 million sequences/lane. 150 million sequences/384 samples = 390,000 sequences/sample.

2<sup>nd</sup> Gene set: 320 genes (x 3 assays/gene) 960 assays being designed

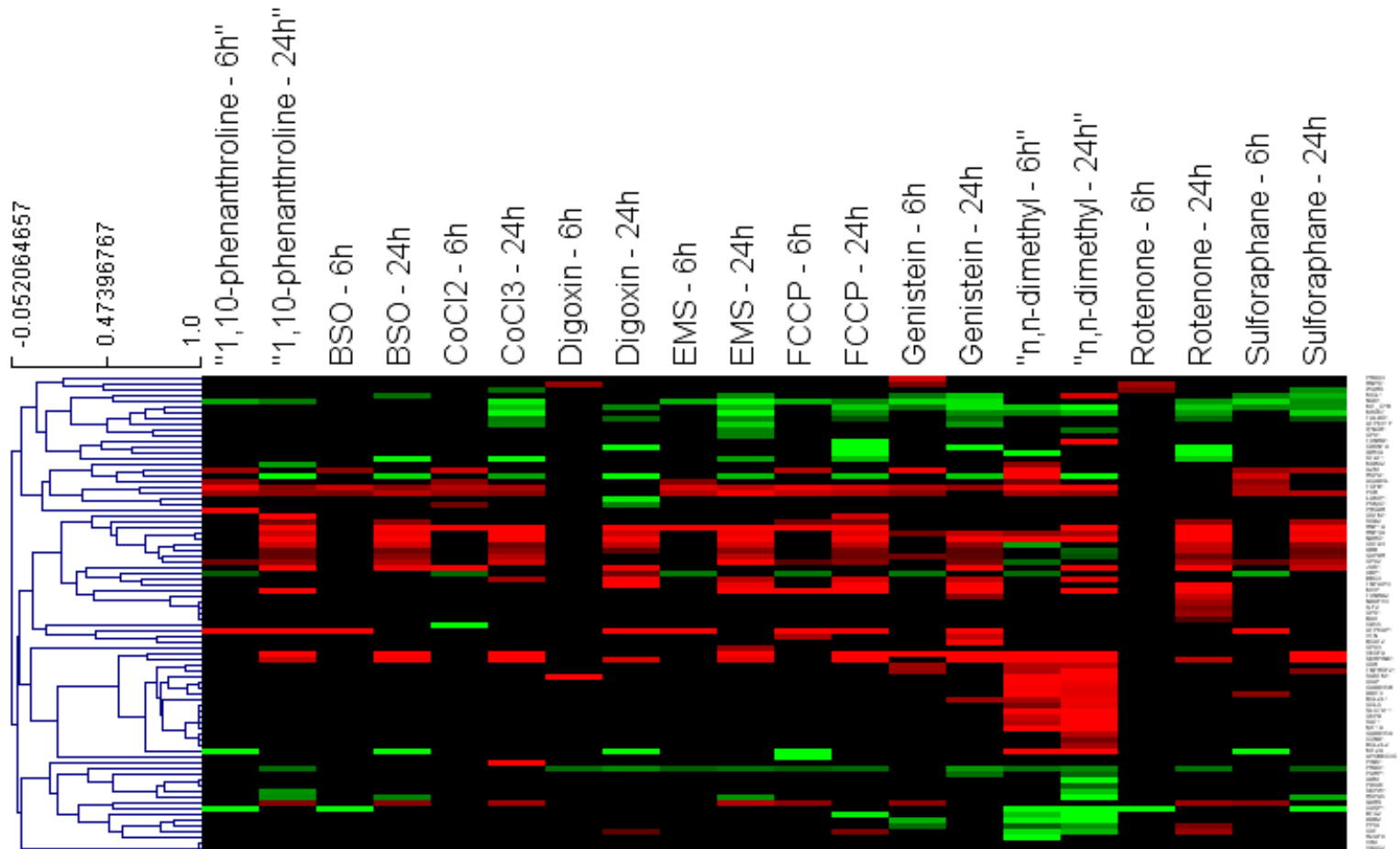
3<sup>rd</sup> Gene set: ~1,000 genes being selected by working group.



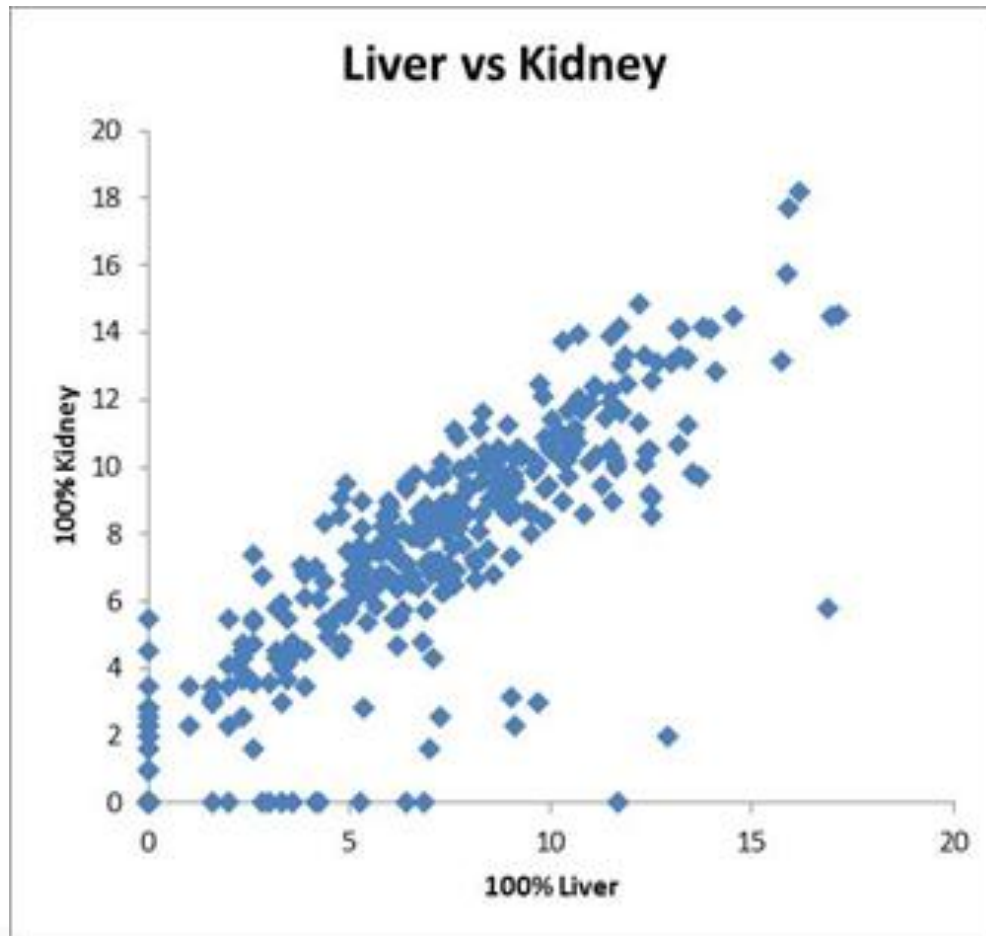


# RASL-Seq Results

50-of-384 samples > 5 million reads on MiSeq

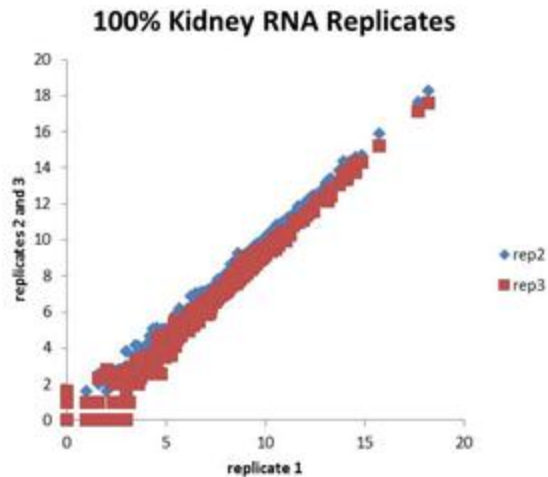
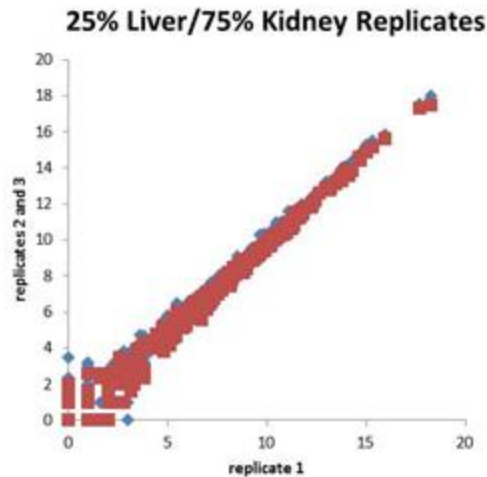
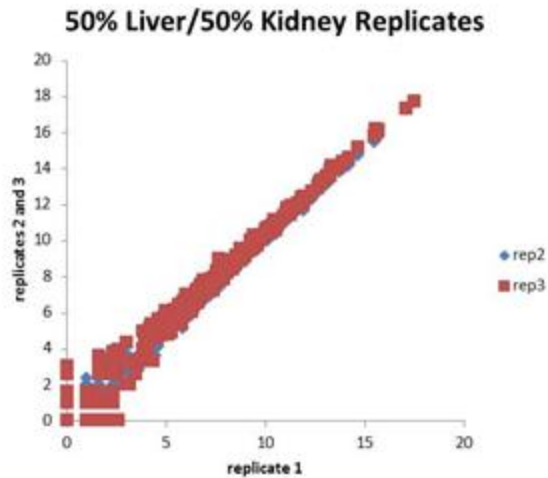
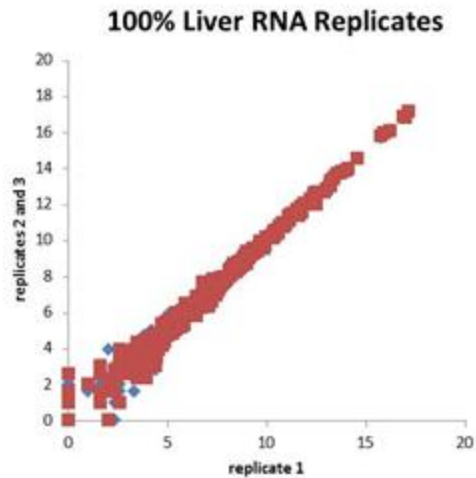


# RASL-Seq Describes Differential Expression in Liver vs Kidney for First 120 Genes



Data courtesy of J. Yeakley, BioSpyder, 14 March 2014

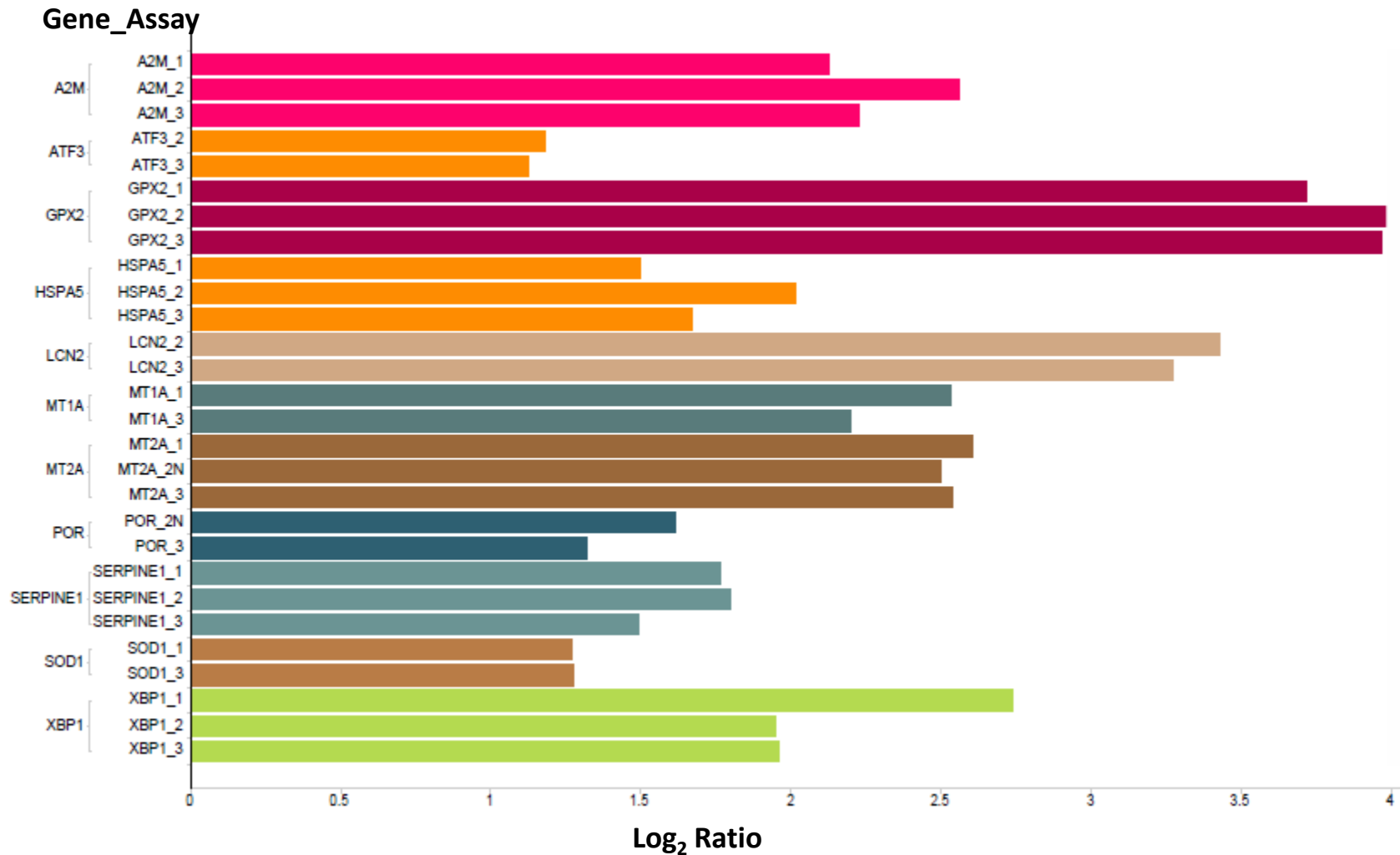
# Intra-Experiment Variability is Low in RASL-Seq



The twelve replicate pairwise  $R^2$  values ranged from 0.990 to 0.995

Data courtesy of J. Yeakley, BioSpyder  
14 March 2014

# Redundant RASL-Seq Assays Give Similar T/C Ratios



# Tox21 Gene Set Selection

Select core set of 1000 genes to assay on all cell lines & samples.

Working Group established to select ~1,000 genes for humans...then other species. Led by Rick Paules

- to: identify ~ 1000 genes that will optimally represent the gene expression responses of the entire genome to diverse chemical and biological challenges.
- We reviewed costs. Selecting new genes and assays will cost ~\$90/gene

# Typical RASL-Seq Application

## Example:

- 60 compounds selected from primary mitochondrial membrane permeability screen Tox21
- Treat 3 models: (*e.g.* HepG2, LUHMES dopa-neurons, cardiomyocytes) x 2 concentrations x 2 time pts x 60 cpds = 720 treatments (2 x 384-well plates x 3 reps?).

## Interpretation:

- Do cpds fall into groups that imply mechanisms?
- Do gene expression changes suggest mechanisms?
- Do cellular models react differently to same cpds?

# Data Analysis & Interpretation

## Data analysis pipeline:

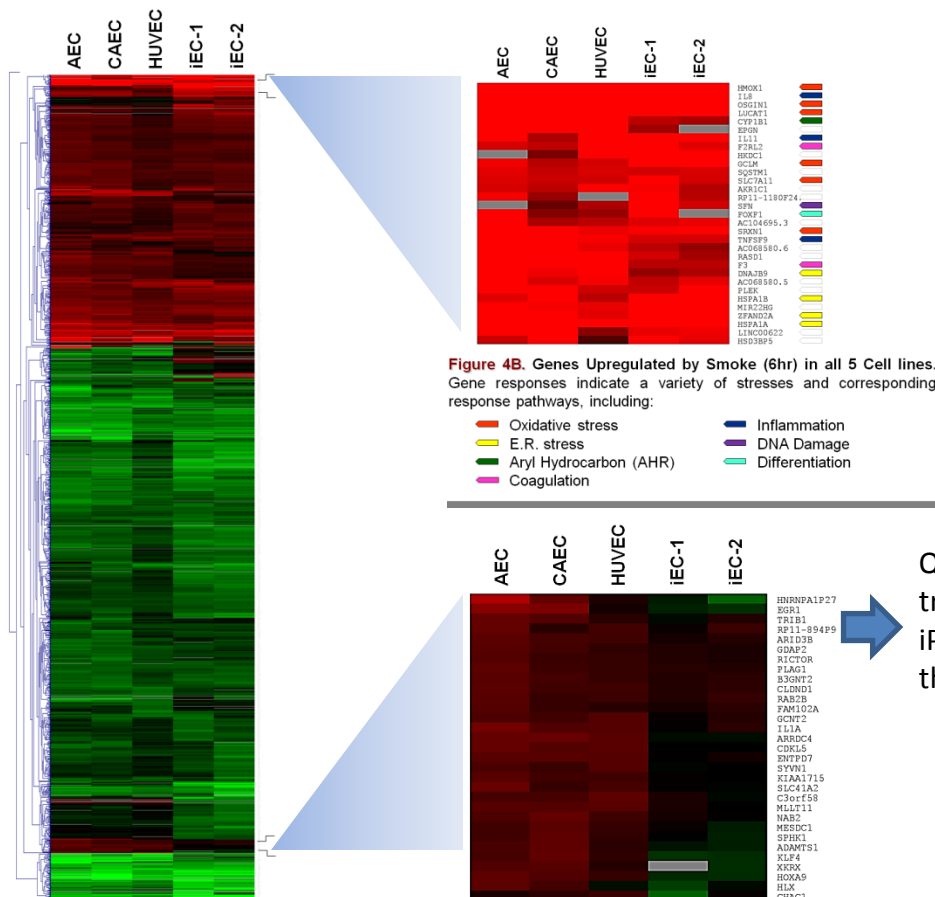
- Filter to remove low quality sequences
- Deconvolute pooled sequences to 384 samples *via* barcodes for rows & columns
- Count matches to target genes (*e.g.* >45/50nt). Adapt NCATS extant pipeline for RNAseq data
- Perform statistical tests for significance for each assay (treated vs controls). Use median if 2-3 assays/gene?

John Braisted's scripts.  
Can be automated

## Data interpretation:

- Associate cpds that cause similar gene expression profiles. Does cpd profile resemble a reference perturbation profile?
  - Implies similar mode or mechanism of toxicity.
- Do early-responding genes *describe* the mode, *e.g.* DNA repair genes or E.R. stress genes?
- Clarify hypotheses by referring to RNAseq baseline data for each cell line. *E.g.* Is the hypothesized receptor/transcription factor/pathway transcribed in these cells?
- Do cellular models react differently to same cpds?

# How Would a Baseline RNAseq Dataset for Each Cell Line Inform RASL-Seq Data? Example:



RNAseq (mRNA) Responses to Tobacco Smoke (6hr) in iPS-derived Endothelial Cells (iECs), and Primary Endothelial Cells

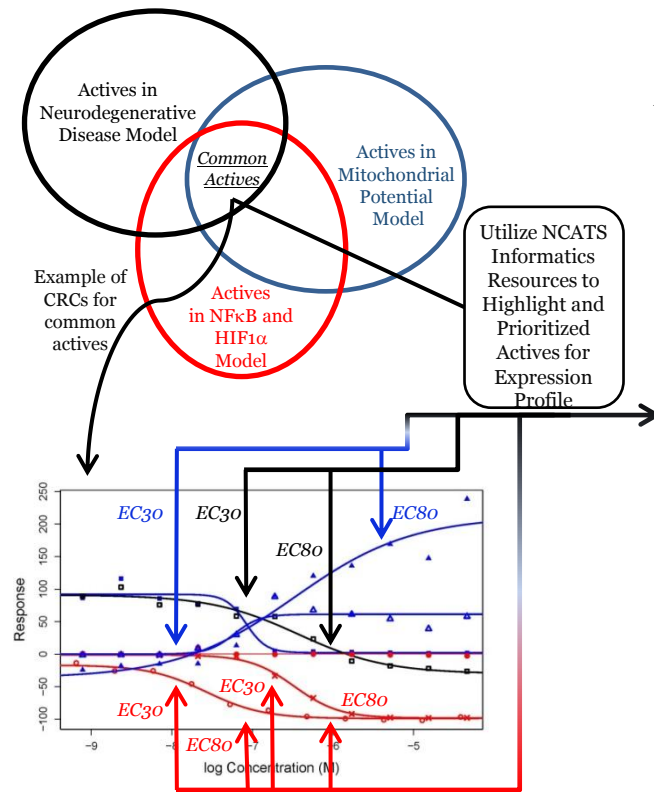
Question: Is a developmental transcription factor missing from the iPS-derived cells that could explain these nonresponsive genes?

Examine reference RNAseq data

RNAseq data suggest GATA6 is expressed only in the primary cells. Testable Hypothesis

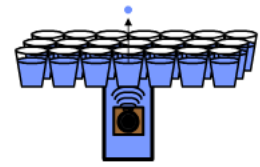


Disease Cell Models	General Phenotype Cell Models	Pathway Informing Cell Models
<b>1) Cancer</b> <ul style="list-style-type: none"> <li>Multiple Myeloma <ul style="list-style-type: none"> <li>Viability &amp; Apoptosis</li> </ul> </li> <li>Rhabdomyosarcoma and Neuroblastoma <ul style="list-style-type: none"> <li>Viability &amp; Apoptosis</li> </ul> </li> </ul> <b>2) Diabetes</b> <ul style="list-style-type: none"> <li>INS-1E pancreatic <math>\beta</math>-cells <ul style="list-style-type: none"> <li>Insulin secretions in</li> </ul> </li> </ul> <b>3) Neurodegenerative Diseases</b> <ul style="list-style-type: none"> <li>Hela line w/GFP-parkin</li> <li>Parkin Translocation</li> </ul> <b>4) Innate Immunity</b> <ul style="list-style-type: none"> <li>A549 line w/GFP-RSV <ul style="list-style-type: none"> <li>RSV spread</li> </ul> </li> <li>Hela line w/GFP-Vaccinia <ul style="list-style-type: none"> <li>Vaccinia spread</li> </ul> </li> </ul>	<b>1) Autophagy</b> <ul style="list-style-type: none"> <li>HEK 293A line</li> </ul> <b>2) Mitochondrial Potential</b> <ul style="list-style-type: none"> <li>Parkin line</li> </ul> <b>3) DNA Damage/Repair</b> <ul style="list-style-type: none"> <li>P53 HCT-116 line</li> </ul> <b>4) TNF<math>\alpha</math> secretion</b> <ul style="list-style-type: none"> <li>THP-1 line</li> </ul> <b>5) ROS Induction/Mitigation</b> <ul style="list-style-type: none"> <li>HEPG2 line</li> </ul> <b>6) Type 1 interferon response</b> <ul style="list-style-type: none"> <li>hFibroblast line</li> </ul> <b>7) Lipid Droplet accumulation</b> <ul style="list-style-type: none"> <li>3T1L line</li> </ul>	<b>1) NF<math>\kappa</math>B</b> <ul style="list-style-type: none"> <li>Me-180 line</li> </ul> <b>2) STAT3</b> <ul style="list-style-type: none"> <li>HEPG2 line</li> </ul> <b>3) HIF1<math>\alpha</math></b> <ul style="list-style-type: none"> <li>ME-180 line</li> </ul> <b>4) NRF2</b> <ul style="list-style-type: none"> <li>HEPG2 line</li> </ul> <b>5) AP1</b> <ul style="list-style-type: none"> <li>ME-180 line</li> </ul> <b>6) Estrogen Receptor</b> <ul style="list-style-type: none"> <li>HEK293 line</li> </ul> <b>7) Androgen Receptor</b> <ul style="list-style-type: none"> <li>HEK293 line</li> </ul> <b>8) CREB</b> <ul style="list-style-type: none"> <li>HEK293 line</li> </ul>



Utilize NCATS Informatics Resources to Highlight and Prioritized Actives for Expression Profile

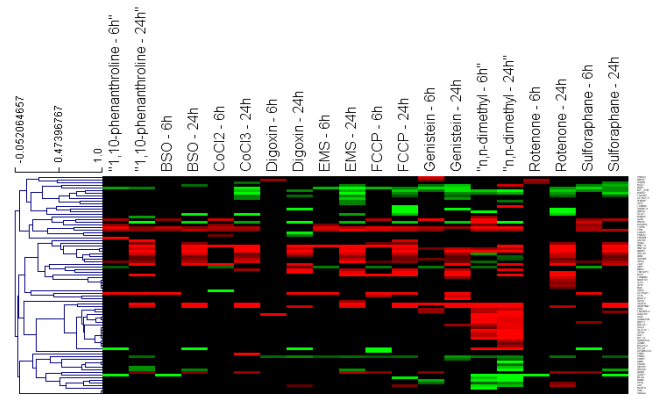
Macros written for EDC acoustic dispenser allow facile plating of each small molecule perturbagen at custom concentrations



Established protocols allow rapid RASL-Seq gene capture on 1000 genes per sample in 384-well plate format

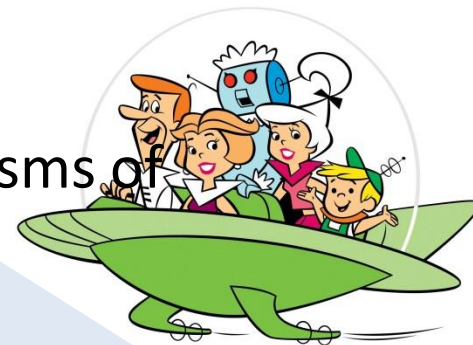


Comparative analysis of expression signatures from selected, active and mirrored inactive perturbagens



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# Collaborators

"Vieles ist bekannt, aber leider in verschiedenen Köpfen", W. Kollath  
("much is known, but unfortunately in different heads")

## NCATS

John Braisted

David Kuo

Pei-Hsuan Chu

David Gerhold

Anton Simeonov

## Tox21 – NTP & EPA

Ray Tice

Rick Paules

## BioSpyder Corp.

JoAnn Yeakley

Joel McComb

Bruce Seligmann

## NHLBI



Manfred Boehm

Avram Walts

## NHLBI Seq Core

Jun Zhu

Yan Luo

Poching Liu

